

BCYE Agar (Legionella Isolation Medium) SKU: 700004412, 700004413, 700004414, 700004415 NCM0037

Intended Use

BCYE Agar is used (with appropriate supplementation) for the isolation of *Legionella* spp. from water and environmental samples. BCYE Agar is not intended for use in the diagnosis of disease or other conditions in humans.

Description

In 1977, McDade *et al.* identified *Legionella pneumophila* as the causative agent of Legionnaires' disease, a multisystem disease manifested primarily by pneumonia. In 1978 a new medium, F-G Agar, resulted in improved growth of *L. pneumophila*, a very fastidious organism. Freely *et al.* modified F-C Agar by substituting yeast extract as a vitamin source and replacing starch with activated charcoal, producing Charcoal Yeast Extract (CYE) Agar. In 1980, Pasculle *et al.* reported that CYE Agar could be improved by the addition of ACES (N-2-acetamido-2-aminoethane sulfonic acid) buffer. One year later, Edelstein further increased the sensitivity of the medium by adding the potassium salt of alpha-ketoglutaric acid.

Typical Formulation

Yeast Extract	10.0 g/L
Charcoal	2.0 g/L
Ferric Pyrophosphate	0.25 g/L
ACES Buffer	10.0 g/L
Potassium Carbonate	2.3g/L
Agar	14.0 g/L

Final pH: 6.9 ± 0.1 at 25°C

Formula is adjusted and/or supplemented as required to meet performance specifications.

Supplements

NCM4007	GVPC Selective Supplement
NCM4006	BCYE Growth Supplement
NCM4005	BCYE Growth Supplement (without L-Cysteine)

Precaution

Refer to SDS

Preparation

1. Suspend 38.5 grams of the medium in 1L of purified water.
2. Heat with frequent agitation and boil for one minute to completely dissolve the medium.
3. Autoclave at 110°C for 10 minutes. Cool to 45 - 50°C.
4. Aseptically add 2 vials of NCM4006-0.5* or 700004869 BCYE Growth Supplement, each reconstituted using 15 mL sterile deionized/RO water, to make BCYE agar with Legionella growth supplements.
5. To make GVPC selective medium additionally add 2 vials of NCM4007-0.5* or 700004871 GVPC Selective Supplement, each reconstituted using 10 mL sterile deionized/RO water.
6. Mix thoroughly and continuing mixing throughout dispensing.

NOTE: NCM4006 supplement can be substituted with NCM4005 supplement (without the addition of NCM4007 selective supplement) to prepare presumptive identification plates. Final pH: 7.1 ± 0.1 at 25°C

Technical Specification Sheet



*Larger vials may be available. Please see appropriate supplement data sheet for availability and preparation instructions

Test Procedure

Methods range from direct inoculation to concentration via membrane filtration with or without pre-treatment. After ensuring inocula has been absorbed invert the plates and incubate at $36 \pm 1^\circ\text{C}$ for 7 to 10 days. Create a humid atmosphere to prevent desiccation of the plates.

Quality Control Specifications

Dehydrated Appearance: Powder is homogeneous, free flowing and black.

Prepared Appearance: Prepared medium is opaque and black.

Expected Cultural Response:

Cultural response on BCYE Agar Supplemented with NCM4006 Growth Supplement at $36 \pm 2^\circ\text{C}$ and examined for growth.

<u>MICROORGANISM</u>	<u>ATCC</u>	<u>APPROX. INOCULUM (CFU)</u>	<u>EXPECTED RESULTS</u>
<i>Legionella bozemanii</i>	33217	50-200	>50% grey/white colonies
<i>Legionella pneumophila</i>	33152	50-200	>50% grey/white colonies
<i>Legionella pneumophila</i>	33156	50-200	>50% grey/white colonies
<i>Pseudomonas aeruginosa</i>	9027	10^3 - 10^4	Growth
<i>Escherichia coli</i>	8739	10^3 - 10^4	Growth
<i>Escherichia coli</i>	25922	10^3 - 10^4	Growth
<i>Enterococcus faecalis</i>	19433	10^3 - 10^4	Growth
<i>Enterococcus faecalis</i>	29212	10^3 - 10^4	Growth

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620 Leshar Place • Lansing, MI 48912
800-234-5333 (USA/Canada) • 517-372-9200
foodsafety@neogen.com • foodsafety.neogen.com

Technical Specification Sheet



Cultural response on BCYE Agar Supplemented with NCM4006 Growth Supplement and NCM4007 GVPC selective supplement at 36 ± 2°C and examined for growth

<u>MICROORGANISM</u>	<u>ATCC</u>	<u>APPROX. INOCULUM (CFU)</u>	<u>EXPECTED RESULTS</u>
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<i>Pseudomonas aeruginosa</i>	9027	10 ³ -10 ⁴	Complete to partial inhibition
<i>Escherichia coli</i>	8739	10 ³ -10 ⁴	Complete to partial inhibition
<i>Escherichia coli</i>	25922	10 ³ -10 ⁴	Complete to partial inhibition
<i>Enterococcus faecalis</i>	19433	10 ³ -10 ⁴	Partial inhibition
<i>Enterococcus faecalis</i>	29212	10 ³ -10 ⁴	Partial inhibition

The organisms listed are the minimum that should be used for quality control testing.

Results

Legionella pneumophila produces small to large, smooth, colorless to pale, blue-grey, slightly mucoid colonies that fluoresce yellow-green under longwave UV light. A gram stain, biochemical tests, and serological procedures should be performed for confirmation of *L. pneumophila*.

Storage

Store sealed bottle containing the dehydrated medium at 2 - 30°C. Once opened and recapped, place container in a low humidity environment at the same storage temperature. Protect from moisture and light by keeping container tightly closed.

Expiration

Refer to expiration date stamped on the container. The dehydrated medium should be discarded if it is not free flowing, or if appearance has changed from the original color. Expiry applies to medium in its intact container when stored as directed.

Limitations of the Procedure

1. Due to nutritional variation, some strains may be encountered that grow poorly or fail to grow on this medium.
2. Biochemical tests and serological procedures must be performed to confirm presence of *L. pneumophila*.



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References

1. McDade, Shepard, Fraser, Tsai, Redus, Dowdle and the Laboratory Investigation Team. 1977. N. Engl. J. Med. 297:1197.
2. Edelstein. 1985. *In* Lennette, Balows, Hausler and Shadomy (eds.). Manual of clinical microbiology, 4th ed. ASM. Washington, D.C.
3. Freely, Gorman, Weaver, Mackel and Smith. 1978. J. Clin. Microbiol. 8:320.
4. Freely, Gibson, Gorman, Lansford, Rasheed, Mackel and Baine. 1979. J. Clin. Microbiol. 10:437.
5. Pasculle, Freely, Gibson, Cordes, Myerowitz, Patton, Gorman, Carmack, Ezzell and Dowling. 1980. J. Infect. Dis. 141:727.
6. Edelstein. 1981. J. Clin. Microbiol. 14:298.